# **PALLADIUM-CATALYSED COUPLING OF VINYL TRIFLATES WITH ENYNES**  AND ITS APPLICATION TO THE SYNTHESIS OF 1a,25-DIHYDROXYVITAMIN **D**<sub>3</sub><sup>1,2</sup>

José L. Mascareñas, Luis A. Sarandeses, Luis Castedo, and Antonio Mouriño\* Departamento de Química Orgánica, Facultad de Química y Sección de Alcaloides del CSIC, 15706 Santiago de Compostela, Spain

#### *(Received* in UK 14 *January* 1991)

**Abstracts.** We describe a general approach, based on the palladium-catalysed coupling of enynes with vinyl triflates, for the construction of dienynes related to vitamin D metabolites and analogues. As an application of this method, an efficient convergent synthesis of  $1\alpha.25$ dihydroxyvitamin D<sub>3</sub> starting from the Inhoffen-Lythgoe diol (6a) and natural carvones has been carried out (11 steps, 28% overall yield from **6a).** This strategy allows labelling of the side chain in the final steps of the synthesis

#### **Introduction**

Vitamin Ds **(la),** before fulfilling classical biological functions such as intestinal calcium absorption and bone calcium mobilization, must undergo hydroxylation at C-25 and C-l to produce 1α,25-dihydroxyvitamin D<sub>3</sub> [1b, 1α,25-(OH)<sub>2</sub>-D<sub>3</sub>]. This metabolite is thought to be the active form of vitamin  $D_3$ , and its mode of action resembles that of the steroid sex hormones.<sup>3</sup> Great interest has recently been attracted in the vitamin D field by the discovery that in addition to playing an important role in the medical treatment of a wide spectrum of human diseases,<sup>3c</sup>  $1\alpha,25-(OH)_2-D_3$  is also able to induce cell differentiation and to inhibit cell proliferation in several lines of cancer cells.4

Several reports of syntheses of  $1\alpha,25$ -(OH)<sub>2</sub>-D<sub>3</sub> have appeared.<sup>5</sup> Most of these approaches follow the long and tedious classical method in which the low-yielding electrocyclic photochemically induced opening of steroidal 5.7-dienes is the key step  $(*18-23$  steps,  $-0.2-1$  % overall yield).<sup>6</sup> More recently, partial syntheses from 25-OH-D<sub>3</sub>7 or from the inexpensive vitamin  $D_2$ <sup>8</sup> and a convergent approach based on the Wittig-Horner coupling of ketones with allylic phosphine oxides,9 have also been reported.

Lythgoe and coworkers have shown that semihydrogenation of dienynes of type 2 (Scheme I), followed by thermolysis of the resulting previtamins, affords the corresponding vitamin D derivatives in good yield.10 However, this approach has three drawbacks: (i) many steps are required for the preparation of the chloroketone corresponding to the CD fragment, (ii) low yield  $(-3\%)$  is obtained in the A ring fragment, and (iii) low yield in dienyne (20 %) is obtained by successive coupling between the chloroketone and an acetylenic fragment corresponding to ring A followed by formal elimination of CIOH.

We have recently reported convenient syntheses of dienynes related to vitamin D metabolites and analogues in which the key step is the palladium-catalysed coupling of enynes and vinyl triflates (Scheme  $1$ ).<sup>2b,11</sup> We now describe details of this procedure and its application to an efficient synthesis of  $1\alpha$ ,25-(OH)<sub>2</sub>-D<sub>3</sub> following the retrosynthetic analysis depicted in Scheme I. This approach also allows labelling of the hydroxylated side chain in the final steps of the synthesis, and is flexible enough to permit preparation of several C-11-substituted analogues of potential interest for (i) the construction of affinity columns for the isolation and purification of the receptor of the hormone, (ii) the construction of photolabile probes for studying the active site of the receptor, and (iii) the generation of specific monoclonal  $1\alpha,25-(OH)_2-D_3$  antibodies.



#### **Syntheses of CD side chain fragments**

**In** preliminary experiments to determine the effectiveness of the coupling reaction between vinyl triflates and the A-ring fragments **4a** and **4b, we** focussed our attention on the simple vinyl triflate **3a** (Scheme I), which had already proved to be a useful model in similar reactions with other A-ring derivatives.11 Compound **3a** was obtained in around 90 % yield (several runs) by reacting Grundmann's ketone (5)12 (Chart I) with lithium di-isopropylamide (LDA) to form the kinetic enolate, followed by quenching with N-phenyltriflimide<sup>13</sup> (Tf<sub>2</sub>NPh).

For the synthesis of the hormone **lb we** chose the vinyl triflate 3b (Scheme I), since the 25 masked carbonyl would allow labelling in the last steps of the synthesis. This compound was efficiently prepared from the Inhoffen-Lythgoe diol **(6a)** (44 %, 6 steps, chart I) as follows: monotosylation of **6a** and treatment of the resulting tosylate **6bl4** with sodium iodide afforded 6c (96 %). The key step for the construction of the side chain was the sonication of a mixture of iodide 6c, methyl vinyl ketone, Cul and Zn in H<sub>2</sub>O:EtOH (3:7) following Luche's method,<sup>15</sup> which provided the known ketoalcohol **7al7** in 76 % yield (this interesting procedure avoids the need to protect the hydroxyl group).16 Ketalization and oxidation of the resulting ketal **7b,** as previously reported, afforded the protected ketone  $7c^{17}$  (72 %). Finally, reaction of 7c with LDA and trapping of the resulting kinetic enolate with Tf<sub>2</sub>NPh gave the desired vinyl triflate 3b (Scheme I) in 90 % yield.

**Chart** I







**5 68**, X=OH **7a**, R<sub>1</sub>=R<sub>2</sub>=O, R<sub>3</sub>=OH, R<sub>4</sub>=H<br>**6b**, X=OTs **7b**, R<sub>1</sub>+R<sub>2</sub>=OCH<sub>2</sub>CH<sub>2</sub>O, R<sub>3</sub>= **6b** , X=OTs **7b** , R<sub>1</sub>+R<sub>2</sub>=OCH<sub>2</sub>CH<sub>2</sub>O, R<sub>3</sub>=OH, R<sub>4</sub>=H **6c** , X=I<br>**7c** , R<sub>1</sub>+R<sub>2</sub>=OCH<sub>2</sub>CH<sub>2</sub>O, R<sub>3</sub>=R<sub>4</sub>=O **7c**, R<sub>1</sub>+R<sub>2</sub>=OCH<sub>2</sub>CH<sub>2</sub>O, R<sub>3</sub>=R<sub>4</sub>=O

# **Syntheses of A-ring fragments**

Diol **4a** and protected diol **4b** (Scheme I) were synthesized either from (R)-(-)-carvone **(8a, I**carvone) or from (S)-(+)-carvone **(8b,** d-carvone) (Chart II). Sodium borohydride reduction of **8a**  under Luche conditions<sup>18</sup> afforded an inseparable mixture of *cis*-carveol<sup>19c</sup> and its *trans-*epimer in a ratio of ca. 12:1 as shown by <sup>1</sup>H NMR analysis (91 %). This mixture was subjected to the Mitsunobu procedure<sup>20</sup> to provide, after column chromatography filtration, the crude benzoate 10a, which without further purification was hydrolysed by means of methanolic potassium hydroxide to a mixture of the desired trans-carveol  $10b^{19b}$  together with approximately 8 % of its epimer, as shown by 1H NMR analysis (70 % yield).

Snags arising during the purification of benzoate **10a** from byproducts, and the expense of the above method, induced us to seek an alternative procedure for the preparation of trans-caweol lob. Stereoselective epoxidation of d-carvone **(8b)** was conveniently accomplished using modified Ohloff conditions (30 % H<sub>2</sub>O<sub>2</sub>, LiOH, MeOH) to give the epoxy ketone  $9^{21,19d}$  in 88 % yield after kugelrohr distillation. Wharton reaction on 9 (NH2NH2\*H2O, Me2NCH2CH2OH)<sup>22</sup> afforded a mixture of *trans*-carveol 10b and the corresponding minor product *cis*-carveol in 60-70 % yield after conventional chromatography and kugelrohr distillation. Gas chromatographic analysis of this mixture showed it to contain 4 % cis-carveol. Potassium hydride deprotonation of alcohol 10b in THF followed by alkylation with  $ICH<sub>2</sub>SnBu<sub>3</sub><sup>23</sup>$  afforded the stannane 10c, which after column chromatography filtration was rapidly treated with n-BuLi in THF to produce the homoallylic alcohol 11a in 72 % yield through [2,3]-sigmatropic rearrangement<sup>24</sup> of the resulting primary carbanion; we were able to accomplish this transformation in similar yield in a one-pot reaction without isolating the stannyl derivative. Hydroxy-directed epoxidation of freshly distilled **11a** under Sharpless' conditions [VO(acac)<sub>2</sub>, TBHP]<sup>25</sup> provided the desired epoxy alcohol **12a** [86] %, <sup>1</sup>H NMR  $\delta$  3.05 (1H, t, J=2.3 Hz, H-6)] and its epoxy diastereomer [7 %, <sup>1</sup>H NMR  $\delta$  3.30 (1H, t, J=2.1 Hz, H-6)], which were separated by flash chromatography26 Oxidative cleavage of **12a**   $(OSO<sub>4</sub>, KIO<sub>4</sub>)$  followed by flash chromatography gave pure 12c in 88 % yield [<sup>1</sup>H NMR  $\delta$  3.08 (1H, t, J=2.2 Hz, H-6)]. At this stage traces of a compound of closely similar Rf, which according to<sup>1</sup>H NMR analysis of the crude (6 2.98, d) made up approximately 4% of the total mass, were also separated. As is argued below, this byproduct was assigned as the enantiomer of **12d,** derived from the diastereomer of the epoxy carvone 9. Conversion of 12c into acetate 12e was then accomplished in 89 % yield by Baeyer-Villiger oxidation (m-CPBA, CHCl3).

In order to check the purity of **12e** and to ensure that no epimerization had taken place at C-4 during the last two steps, cis-carveol **IOd** obtained from d-carvone (8b) under Luche's conditions was subjected to the same sequence of reactions as above (10d $\rightarrow$ 10e $\rightarrow$ 11b $\rightarrow$ 12b $\rightarrow$ 12d $\rightarrow$ 12f) to afford acetate **12f** in 30 % overall yield. The <sup>1</sup>H NMR of **12d** shows a doublet centred at  $\delta$  2.97 identical with that observable as a small signal in the <sup>1</sup>H NMR spectrum of crude 12c (see above). In view of the synthetic sequence, this suggests that the material separated as traces from 12c is the enantiomer of **12d.** 

Swern oxidation28 of **128** under appropriate conditions opened the epoxy ring to afford the hydroxy acetate **13a,** which was acetylated to the diacetate **13blOa** in 83 % yield (two steps). Chain extension<sup>29</sup> to diol 4a (Scheme I) was accomplished in 73 % yield by conversion to the vinyl dibromide **14a** (Zn, CBr4, PhsP, pyridine) and subsequent elimination with n-BuLi. Standard protection with tert-butyldimethylsilyl chloride afforded the desired enyne 4b<sup>30</sup> (16 % overall yield from d-cawone, 12 steps).



Alternatively, it was also possible to obtain the enyne **4b** from the hydroxy acetate **13a as**  follows. Sodium methoxide saponification of **13a** to the hydroxy aldehyde 13c,<sup>31</sup> followed by protection with tert-butyldimethylsilyl chloride, afforded 13d<sup>31,32</sup> in 82 % yield. Chain extension to vinyl dibromide **14b** and subsequent elimination as above afforded **4b** (Scheme I) in 84 % yield (16.5 %, 12 steps from d-carvone).

# **Coupling Experiments**

We first investigated the synthesis of unprotected dienyne **2a** starting from the enyne diol **4a**  and the simple vinyl triflate **3a** (Scheme I). The coupling reaction between **3a** and **4a** in the presence of a catalytic amount of bis(triphenylphosphine)palladium (II) chloride  $[(Ph_3P)_2PdCl_2]$ under previously reported conditions (NEt<sub>3</sub>, DMF, 75 °C)<sup>34</sup> afforded the dienyne diol 2a in 54% yield. This unprotected dienyne was fairly unstable, and for this reason and in view of published precedents,13 we decided to prepare the protected dienyne **2b,** the precursor of the natural hormone **1 b,** starting from the protected enyne **4b.** Coupling between the tert-butyldimethylsilylprotected A-ring fragment **4b** and the 25-carbonyl-protected vinyl triflate **3b** under the above reaction conditions furnished the desired dienyne **2b** in 91% yield after chromatography.

# Synthesis of  $1\alpha$ , 25-dihydroxyvitamin  $D_3$  (Scheme II)

Semihydrogenation of dienyne **2b** in hexane with Lindlar palladium catalyst poisoned with quinoline, with careful monitoring of the reaction by TLC to avoid over-reduction of the triple bond, afforded **15** in 95 % yield after flash chromatography. The previtamin **15** was refluxed in iso-octane for 4 h to bring about equilibration to the vitamin 16. Flash chromatograpy of the crude product afforded a mixture of 15 and 16 (94 % combined yield) in a ratio of approximately 12:88 (<sup>1</sup>H NMR analysis). Treatment of this previtamin-vitamin mixture with H+ cation exchange resin in methanol followed by flash chromatography afforded the pure keto vitamin 17 (81 % yield from 16). Finally,

reaction of 17 with methyllithium in diethyl ether provided the desired  $1\alpha$ , 25-dihydroxyvitamin D<sub>3</sub> metabolite **1 b** in 80 % yield [28 % overall yield from the Inhoffen-Lythgoe diol **(8a)].** 



### **Experimental Section**

General. All dry solvents were distilled under argon. Diethyl ether and tetrahydrofuran (THF) were distilled from sodium/benzophenone. Methylene chloride was distilled from phosphorus pentoxide. Pyridine was distilled from potassium hydroxyde. Di-isopropylamine from Aldrich was distilled and then redistilled from calcium hydride. Dimethylformamide (DMF, from Merck) was stored under Type 4A molecular sieves. All reactions were conducted under an argon atmosphere unless otherwise stated. Boiling points and melting points (open capillary tubes) are uncorrected. Kugelrohr distillation boiling points refer to the external air bath temperature. All 250 MHz <sup>1</sup>H NMR spectra were recorded in a Bruker WM-250 apparatus, using CDCl<sub>3</sub> as solvent. Chemical shifts are given in  $\delta$  units with respect to tetramethylsilane as internal standard. Mass spectra were measured in a Kratos MS-50 apparatus, using electron impact at 70 eV. Mass spectra data are given as m/z (intensity expressed as percentage of total ion current). Flash chromatography was performed by Still's method.<sup>35</sup> Sonications were carried out in a BANDELIN type RK102H apparatus.

(1S,4S,6S)-1-Methyl-4-(1-methylethenyl)-7-oxablcyclo (4.1.0) heptan-2-one (9). A mixture of LiOH.H<sub>2</sub>O  $(25\%$ , 16 mL) and methanol  $(500 \text{ ml})$  was stirred for 5 min.  $(S)-(+)$ -Carvone  $(8b, 9.6 \text{ q}, 64 \text{ mmol}, 10 \text{ ml})$  and hydrogen peroxide (30%, 100 mL, 0.96 mol) were successively added at 0 °C, and the resulting mixture was stirred for 2.5 h and then poured into water (60 mL) and extracted with CH2Cl2 (3 x 70 mL). The combined organic extracts were washed with brine and then dried over Na2S04. After filtration, the solvent was evaporated and the crude residue was bulb to bulb distilled to afford 9.3 g of the epoxide  $9^{21,19d}$  (88 %, bp 70 °C / 0.6 mm) as a colourless oil. <sup>1</sup>H NMR  $\delta$  1.40 (3 H, s, C<sub>1</sub>-CH3), 1.72 (3 H, s, vinyl CH3), 1.7-2.2 and 2.3-2.8 (5 H, two m), 3.45 (1 H, dd, J= 0.9 and 3 Hz, H-6), 4.72 and 4.80 (2 H, two br s, vinyl H's).

(1S,5R)\_2-Methyl-5-(1-methylethenyl)-cyclohex-2-en-l-ol (lob). A mixture of the epoxy ketone 9 (5.5 g, 33.1 mm@), N,N-dimethylethanolamine (15 mL) and NH2NH2.H20 (1 mL, 0.93 g, 18.5 mmol) was stirred at room temperature for 12 h. More hydrazine (1 mL) was added and the resulting mixture stirred for 10 h. Concentration with a rotary evaporator and purification of the residue through a short column of silica gel (10% EtOAc/hexanes) gave 3.4 g of the allyl alcohol 10b<sup>19b</sup> (70 %, colourless viscous liquid), which was further purified by kugelrohr distillation (bp 80 °C/0.1) mm). <sup>1</sup>H NMR  $\delta$  1.75 (3 H, br s, vinyl CH<sub>3</sub>), 1.80 (3 H, br s, C<sub>2</sub>-CH<sub>3</sub>), 4.02 (1 H, m, H-1), 4.73 (2 H, m, vinyl H's), 5.60 (1 H, m, H-3). A weak signal at 6 4.19 (approx. 4 % intensity relative to the signal at 4.02) was assigned to H-l of the epimer at C-l. G. C. analysis (Column: 6 ft, 3 % OV-17. 170 °C) of the distilled reaction product showed the same isomer ratio.

(1R,5R)-2-Methyl-5-(1-methylethenyl)-cyclohex-2-en-1-ol [(-)-cis-carveol, epimer of 10b at C-11. (R)-(-)-carvone (6a,lO g, 66.6 mmol) and CeCl3.7H20 (25.3 g, 68 mmol) were dissolved in dry MeOH (100 mL). The solution was cooled to 0 °C, and NaBH<sub>4</sub> (2.5 g, 68 mmol) was added in portions. After stirring for 30 min at room temperature, water was carefully added and the resulting mixture was extracted with  $E_2O$  (3 x 100 mL). The combined organic layers were washed with brine (50 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and filtered. Concentration in the rotary evaporator afforded 9.1 g of crude epimer<sup>19c</sup> of 10b, which was subjected to the Mitsunobu inversion. <sup>1</sup>H NMR  $\delta$  1.74 and 1.76 (6 H, two br s, vinyl CH<sub>3</sub>'s), 4.19 (1 H, m, H-l), 4.73 (2 H, br s, vinyl H's), 5.50 (1 H, m, H-3). The weak signal at 6 4.02 (approx. 8 % intensity relative to the signal at 4.19) is attributed to H-1 of 10b.

(1S,5R)-2-Methyl-5-(1-methylethenyl)-cyclohex-2-en-1-ol (10b). To a stirred mixture of the above crude ciscarveol (1g, 6.6 mmol), PhCO<sub>2</sub>H (1.6 g, 13.1 mmol) and Ph<sub>3</sub>P (3.5 g, 13.2 mmol, crystallized from hexane) in THF (70 mL) was slowly added a solution of diethylazodicarboxylate (2.1 mL, 2.3 g, 13.1 mmol) in THF (13 mL). The mixture was stirred for 15 h, topped up with more Ph<sub>3</sub>P (3.5 g, 13.2 mmol), PhCO<sub>2</sub>H (1.6 g, 13.1 mmol) and diethylazodicarboxylate (2.1 mL), and stirred for an additional 20 h. The mixture was rotary evaporated to a small volume, diluted with hexane/Et2O (1:1) and washed with brine. The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated under reduced pressure. The resulting residue was filtered through a column of silica gel (2% EtOAc/hexanes) to give 1.23 g of the epimeric benzoate  $(72%)$ , which was subjected to the next reaction.  $1H NMR \delta 1.74$  and 1.75 (6H, two br s, vinyl CH<sub>3</sub>'s), 4.73 (2H, two br s, vinyl H's), 5.51 (IH, m, H-3), 5.81 (IH, m, H-l), 7.40-7.60 (5H, m, Ar).

To a solution of the benzoate obtained as above in MeOH (IO ml) was added KOH (28 g, 0.7 mol) dissolved in MeOH (200 mL). The mixture was stirred for 14 h and the solvent removed in vacuo. The residue was diluted in Et<sub>2</sub>O (150 mL) and the resulting solution washed with brine. The aqueous layer was extracted again with Et<sub>2</sub>O (90 mL). The combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in vacuo. The residue was purified through a colummn of silical gel (5% EtOAc/hexanes) to give 3.1 g of 10b (95%), whose <sup>1</sup>H NMR spectrum was similar to that of the product obtained previously, although now there was approx. 8% contamination by the epimer.

Bu<sub>3</sub>SnCH<sub>2</sub>I. Zn-Cu couple prepared by Smith's<sup>36</sup> procedure (10 g) and dry THF (20 mL) were placed in a three-necked round-bottomed flask equipped with a thermometer, a dropping funnel and a reflux condenser. CH<sub>2</sub>I<sub>2</sub> (4.91 mL,16 g, 59.6 mmol) was dissolved in THF (14 mL) in the dropping funnel. The reaction was iriitiated by the addition of a few drops of the CH<sub>2</sub>1<sub>2</sub> solution, and then this solution was added dropwise at a rate which maintained the temperature between 38 and 40 °C. The mixture was stirred for 1.5-2 h, cooled in an ice bath, and filtered through a Schlenk-type apparatus into another dry three-necked flask equipped with a thermometer and dropping funnel. To the white filtrate, a solution of freshly distilled BugSnCl (16 mL, 21 g, 64 mmol) in THF (20 mL) was added dropwise over 30 min from the dropping funnel. After stirring for 12 h, the reaction mixture was poured into an aqueous solution of HCI (10 %, !OO mL) and extracted with hexanes. The organic layer was washed with brine, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator to give a residue which was filtered through a column of silica gel (hexanes) to afford liquid Bu<sub>3</sub>SnCH<sub>2</sub>I,<sup>23</sup> which was further purified by kugelrohr distillation [24 g, 93 %, bp 110-120 °C/0.1 mm; Rf: 0.90 (hexanes)]. <sup>1</sup>H NMR  $\delta$  1.94 (2 H, s, -CH2Sn-).

(1R,5R)-2-Methyl-5-(1-methylethenyl)-2-cyclohexen-l-methanol (1 la). An excess of KH (35 % dispersion in mineral oil) was washed with hexanes (3 x 20 mL) and THF (15 mL) by magnetic stirring and decantation, and dried in high vacuum. To a stirred suspension of dried KH (2.1 g, 52 mmol) in THF (60 mL) was added a solution of 10b (4.6 g, 30 mmol, freshly distilled in the kugelrohr) in THF (26 ml). After stirring at room temperature for 5 h. the mixture was cooled at 0 "C and BugSnCH2I (15 mL, 21.5 g, 50 mmol, freshly distilled in the kugelrohr) was added. The reaction mixture was stirred for 12 h. and MeOH (3 mL) and water (50 mL) were added. The resulting solution was concentrated in the rotary evaporator to a small volume. Extraction with hexanes (3 x 75 mL), washing with brine, drying (Na<sub>2</sub>SO<sub>4</sub>) and concentration in the rotary evaporator afforded an oily residue which was filtered through a column of silica gel (hexanes) to give 12.4 g of stannane 10c (90 %, colourless oil), which was inmediately subjected to the next reaction.<sup>1</sup>H NMR  $\delta$  1.75 (6 H, br s, vinyl CH3's), 3.36 (1 H, br s, H-l), 3.55 and 3.95 (2 H. AB, J= 10 Hz, -OCH2Sn-), 4.73 (2 H, m, vinyl H's), 5.54 (1 H, m, H-3). To a stirred solution of stannane 10c (12.4 g, 27.2 mmol) in THF (80 mL) at -78 °C was added a solution of n-butyllithium i hexane (17.5 ml, 2.1 M, 36.5 mrnol). The reaction mixture was allowed to come to room temperature and was stirred for an additional hour. Several drops of water were added and the solvent removed in the rotary evaporator. The residue was diluted with Et<sub>2</sub>O (100 mL) and the organic layer washed with brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and filtered. Concentration in the rotary evaporator and flash chromatography (5-10 % EtOAc/hexanes) afforded, after concentration in vacuo and high vacuum drying, 3.58 g of the hornoallyl alcohol 11a:  $[a]^{25}D -18.5$ ° (c 0.77, CHCl3); (80 %, 90-100 °C/0.5 mm, viscous colourless liquid). <sup>1</sup>H NMR  $\delta$  1.72 and 1.74 (6 H, two br s, vinyl CH3's), 3.61 (1 H, dd, J= 8 and 11 Hz, -CHHOH), 3.74 (1 H, dd, J= 11 and 3.5 Hz, -CHHOH), 4.73 (2 H, br s, vinyl H's), 5.56 (1 H, m, H-3); MS, m/z 166 (M+, 7), 151 (M+-CH3, 6), 148 (M<sup>+</sup>-H<sub>2</sub>O, 9), 135 (15), 133 (17), 119 (11), 107 (40), 105 (26), 93 (100), 91 (41); HRMS calcd for C<sub>11</sub>H<sub>18</sub>O 166.1353, found 166.1354. 11b:  $[\alpha]^{25}D +11.4^{\circ}$  (c 0.65, CHCl<sub>3</sub>).

(1R,2S,4R,6S)-2-Hydroxymethyl-l-methyl-4-(me~hylethenyl)-7-oxablcyclo [4.1.0] heptane (12a). To a solution of the freshly kugelrohr distilled alcohol 118 (1.55 g, 9.3 mmol) in dry benzene (75 ml) was added vanadyl acetyl acetonate (100 mg), and the resulting mixture was stirred at 50 °C for 5 min. After cooling to room temperature, the reaction mixture was treated dropwise with a solution of anhydrous fert-butylhydroperoxide in toluene (4.3 mL, 3 M, 13 mrnol). The green solution turned deep red as the t-BuOOH was added. The reaction was monitored by TLC (30 % EtOAc/hexanes) and judged complete after 3 h, by which time the deep red colour had turned to yellow. The reaction mixture was poured on a saturated solution of NaHCO<sub>3</sub> (70 mL) and extracted with Et<sub>2</sub>O (3 x 60 mL). The combined organic extracts were dried (MgSO4). filtered and concentrated in the rotary evaporator, and the residue **was** purified by flash chromatography (10-15 %, EtOAc/hexanes) to afford, after concentration in vacuo and high vacuum drying, 1.45 g of the desired 128 (86 %) and 130 mg of the epoxydiastereomer (7 %, less polar compound) as cotourtess oils. Compound 12a : [ $\alpha$ ]<sup>25</sup><sub>D</sub> - 6.4°, (*c* 1.08, CHCl<sub>3</sub>);<sup>1</sup>H NMR  $\delta$  1.40 (3 H, s, C<sub>1</sub>-CH<sub>3</sub>), 1.71 (3 H, br s, vinyl CH<sub>3</sub>), 3.05 (1 H, t, J= 2.3 Hz, H-6), 3.82 (2 H, d, J≖ 5.8 Hz, -CH<sub>2</sub>OH), 4.68 and 4.74 (2 H, two s, vinyl H's); IR (CHCl<sub>3</sub>) 1450, 1370, 1040, 900 cm<sup>-</sup> '; MS, m/z 182 (Mf 2), 167 (M+-CH3,4), 164 (M+-H20,8) 149 (M+-CH3-H2C, 14) 133 (20) 121 (23), 109 (30). 107 (46) 105 (27), 93 (49), 83 (52), 67 (83), 55 (59), 43 (100). HRMS calcd for C<sub>11</sub>H<sub>18</sub>O<sub>2</sub> 182.1302, found 182.1303. Epoxydiastereomer of 12a <sup>1</sup>H NMR  $\delta$  1.44 (3 H, s, C<sub>1</sub>-CH<sub>3</sub>), 1.71 (3 H, br s, vinyl CH<sub>3</sub>), 3.30 (1 H, t, J= 2.1 Hz, H-6), 3.91 (2 H, br s, -CH<sub>2</sub>OH), 4.68 and 4.75 (2 H, two s, vinyl H's). 12b: [ $\alpha$ ]<sup>25</sup>D + 4.2°, (c 2.5, CHCl<sub>3</sub>).

(1R,2S,4R,6S)-2-Hydroxymethyl-4-acetyl-l-methyl-7-oxablcyclo [4.1.0] heptane (12~). To a mixture of 12a (2.15 g, 11.8 rnol) in THF-water (1 :l, 160 mL) were added an aqueous solution of osmium tetroxide (1 %, 2.3 mL) and finely powdered potassium periodate (6.9 g, 30 mmol). After stirring at room temperature for 3 h, additional amounts of OsO<sub>4</sub> (1 %, 1 mL) and KIO<sub>4</sub> (2 g, 8 mmol) were added and the reaction mixture was stirred for a further 12 h. The resulting solution was concentrated in the rotary evaporator, and the residue was diluted with brine (75 mL) and extracted with EtOAc (4 x 50 mL). The combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator, and the resulting residue was purified by flash chromatography (30-50 % EtOAc/hexanes) to afford, after concentration in vacuo and high vacuum drying,1.9 g of the pure ketone 12c (88 %, colourless oil) and traces of a compound of slightly smaller Rf for which we attribute the structure of the enantiomer of 12d, on the basis of caracteristic signals observed in the <sup>1</sup>H NMR of the reaction crude. Compound 12c, <sup>1</sup>H NMR  $\delta$  1.40 (3 H, s, C<sub>1</sub>-CH<sub>3</sub>), 2.16 (3 H, s, CH3CO), 2.64-2.77 (1 H, m, H-4), 3.08 (1 H, 1, J= 2.2 Hz, H-6). 3.77-3.90 (2 H, m, -CH20H); IR (CHC13) 3400 (br), 1700, 1350, 1040 cm-'; MS, nVz 164 (M+, 40) 169 (M+-CH3, 18) 166 (M+-H20. 64), 155 (60), 153 (85), 151 (100) 148 (72); HRMS calcd for C<sub>10</sub>H<sub>16</sub>O<sub>3</sub> 184.1116, found 184.1107. Enantiomer of 12d : <sup>1</sup>H NMR  $\delta$  2.98 (d).

(1R,2S,4R,6S)-2-Hydroxymethyl-4-acetoxy-l-methyl-7-oxablcyclo 14.1.01 heptane (126). Purified *m-*CPBA (2.6 g, 15.1 mmol) was added to an ice cooled solution of ketone 12c (1.6 g, 8.7 mmol) in CHCl<sub>3</sub> (80 mL). An aqueous buffer of pH 7.8 (NaH<sub>2</sub>PO<sub>4</sub>-Na<sub>2</sub>HPO<sub>4</sub>, 70 mL) was added and the resulting mixture stirred for 3h. An additional amount of m-CPBA (2.7 g, 15.7 mmol) was added. After further stirring for 22 h, the reaction mixture was transferred into a separating funnel containing a solution of NaHCO<sub>3</sub> (60 mL). The aqueous layer was extracted with CHCl<sub>3</sub> (3 x 70 mL) and the combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>) and filtered. Concentration in the rotary evaporator afforded a residue which was purified by flash chromatography (70 %, EtOAc/hexanes) to give, after concentration in vacuo and high vacuum drying, 1.55 g of acetate 12e (89 %, colourless oil). <sup>1</sup>H NMR  $\delta$  1.44 (3 H, s, C<sub>1</sub>-CH<sub>3</sub>), 2.04 (3 H, s, AcO), 2.96 (1 H, d, J= 4 Hz, H-6), 3.80 (1 H, dd, J= 4.4 and 11 .O Hz, -CHHOH), 3.95 (1 H, dd, J= 5.2 and 11 .O Hz, CHHOH), 4.96 (1 H, m, H-4); IR (CHCl3) 3000, 1730, 1260,1050 cm<sup>-1</sup>. MS, m/z 182 (M<sup>+</sup>-H<sub>2</sub>O, 1), 157 (M<sup>+</sup>-CH<sub>3</sub>CO, 12), 141 (13), 140 (100), 125 (35), 122 (88), 110 (33), 109 (58), 95 (51), 84 (32), 82 (90). HRMS calcd for C<sub>10</sub>H<sub>16</sub>O<sub>4</sub> -CH<sub>3</sub>CO 157.0885, found i 57.0874.

(3S,5R)-5-Acetoxy-3-hydroxy-2-methylcyclohex-l-ene-l-carboxaldehyde (13a). To a stirred solution of oxalyl chloride (0.164 mL, 1.9 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) at -60 °C was added Me<sub>2</sub>SO (0.27 mL, 3.8 mmol). The mixture was stirred for 5 min and then a solution of alcohol 12e (300 mg, 1.5 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added within 5 min; stirring was continued for an additional 15 min. TEA (2.1 mL, 15 mmol) was added and the reaction was allowed to reach 0-10 °C. After stirring for 5 h at that temperature, water (30 mL) was added and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (40 mL). The organic layer was washed with brine. The aqueous layer was re-extracted with CH2Cl2 (3 x 50 ml). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator. The residue was flash chromatographed (25-35 % EtOAc/hexanes) to afford, after concentration in vacua and high vacuum drying, 260 mg of

aldehyde 13a (88 %, light yellow oil). <sup>1</sup>H NMR  $\delta$  2.04 (3 H, s, Ac), 2.30 (3 H, s, C<sub>2</sub>-CH<sub>3</sub>), 4.38 (1 H, t, J= 3.8 Hz, H-3), 5.18 (1 H, m, H-5), 10.16 (1 H, s, HCO); IR (Film) 3440 (br), 2940, 2870, 1740, 1680, 1250 cm<sup>-1</sup>; MS m/z 198 (M<sup>+</sup>, 1), 168 (1), 155 (1), 152 (8), 136 (7), 135 (50), 134 (100), 119 (80), 105 (95).

(3S,5R)-3,5-Diacetoxy-2-methylcyclohex-1-en-1-carboxaldehyde (13b). Acetic anhydride (0.083 mL, 82 mg, 0.8 mmol, freshly distilled) and dimethylaminopyridine (50 mg) were successively added to a solution of alcohol 13a (100 mg, 0,5 mmol) in pyridine (2 mL). The mixture was stirred at 25 °C for 6 h and then diluted with EtOAc (50 mL). The resulting solution was washed with brine, dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator to give 115 mg of 13b<sup>10b</sup> (95 %, yellow oil), which was used directly in the next step. <sup>1</sup>H NMR δ 2.05 (3 H, s, C<sub>5</sub>-Ac), 2.14 (3 H, s, C<sub>3</sub>-Ac), 2.15 (3 H, s, C<sub>2</sub>-CH<sub>3</sub>), 5.13 (1 H, m, H-5), 5.57 (1 H, t, J= 4 Hz, H-3), 10.17 (1 H, s, HCO); IR (film) 2940,1735,1720, 1670, 1370, 1230, 1040 cm<sup>-1</sup>; MS, m/z 211 (M<sup>+</sup>-HCO, 1), 198 (10), 197 (M<sup>+</sup>-CH3CO, 100), 180 (15), 179 (20), 168 (8), 155 (90).

(3S,5R)-3,5-Dihydroxy-1-ethynyl-2-methylcyclohex-1-ene (4a). To a mixture of zinc dust (192 mg, 2.94 mmol) and Ph<sub>3</sub>P (764 mg, 2.94 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) at room temperature was added CBr<sub>4</sub> (960 mg, 2.94 mmol), and the resulting mixture was stirred for 15 min. Pyridine (0.4 mL) and aldehyde 13b (115 mg, 0.48 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (4 mL) were successively added. After stirring for 30 min, Et<sub>2</sub>O (20 mL) was added and the resulting suspension filtered through a short pad of silica gel [Et2O (2 x 25 mL) and EtOAc (20 mL)]. Concentration of the filtrate in the rotary evaporator afforded the crude vinyl dibromide 14a, which was used in the next step without further purification. <sup>1</sup>H NMR  $\delta$  1.66 (3 H, s C<sub>2</sub>-CH<sub>3</sub>), 2.05 (3H, s, C<sub>5</sub>-Ac), 2.09 (3H, s, C<sub>3</sub>-Ac), 5.03-5.17 (1H, m, H-5), 5.42 (1H, t, J= 3.5Hz, H-3), 6.94 (1H, s, vinyl H). MS, m/z 396 (M+, 1), 394 (2), 392 (1), 337 (30), 335 (50), 333 (30), 313 (10),311 (20), 309 (10), 396 (70), 294 (100), 292 (70).

The crude product of the previous reaction was dissolved in dry THF (6 mL) and cooled to -78 °C under an atmosphere of argon. A solution of n-butyllithium in hexane (1.6 mL, 2.1 M, 3.5 mmol) was slowly added to a solution of the dibromide 14a in THF (6 mL) at -78 °C. The resulting deep red mixture was warmed to room temperature and quenched with a saturated aqueous solution of NH4Cl (5 mL). The resulting mixture was extracted with EtOAc (3 x 20 mL), and the combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator to give a residue which was purified by flash chromatography (80 % EtOAc/hexanes) to give 53 mg of the enynediol 4a (73 % yield from 13b), as a viscous oil that decomposed on standing at room temperature. <sup>1</sup>H NMR  $\delta$  2.01 (3 H, br s, 1/2 W= 12 Hz, C<sub>2</sub>-CH<sub>3</sub>), 3.10 (1 H, s, sp-CH), 4.13 (1 H, m, H-5), 4.25 (1 H, t, J=4.3 Hz, H-3).

(3S,5R)-3,5-Bis[(tert-butyldimethylsilyl)oxy]-2-methylcyclohex-1-en-1-carboxaldehyde (13d). The hydroxyacetate 13a (480 mg, 2.42 mmol) was dissolved in MeOH (4 mL) and the solution was cooled to 0 °C and treated with a solution of sodium methoxide in MeOH (14 mL, 0.2 M, 2.80 mmol). After stirring for 5 h at room temperature, acetic acid (0.5 mL) was added. Concentration in the rotary evaporator gave a residue which was purified by flash chromatography (80 % EtOAc/hexanes) to afford, after concentration in vacuo and high vacuum drying, 300 mg of deprotected diol 13c<sup>31</sup> (90 %, oil). <sup>1</sup>H-NMR  $\delta$  2.27 (3 H, br s, C<sub>2</sub>-CH<sub>3</sub>), 4.17 (1 H, m, H-5), 4.40 (1 H, t, J= 3.5 Hz, H-3), 10.17 (1 H, s, HCO).

A solution of this diol in CH<sub>2</sub>CI<sub>2</sub> (10 mL) was treated with tert-butyldimethylsilyl chloride (610 mg, 4.03 mmol) and imidazole (580 mg, 8.4 mmol). After stirring at room temperature for 20 min, additional amounts of tert-butyldimethylsilyl chloride (300 mg, 2 mmol) and imidazole (280 mg, 4 mmol) were added. The reaction mixture was stirred for 10 h, and then quenched by the addition of water (30 mL). The resulting mixture was extracted with CH<sub>2</sub>CI<sub>2</sub> (2 x 30 mL), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator to gave a residue which was purified by flash chromatography (0-5 %

EtOAc/hexanes) to afford 720 mg of the protected aldehyde 13d<sup>31,32</sup> (90 %, syrup). <sup>1</sup>H NMR  $\delta$  0.05 and 0.07 (6 H, two s, Me<sub>2</sub>Si), 0.13 and 0.15 (6 H, two s, Me<sub>2</sub>Si), 0.87 (9 H, s, Me<sub>3</sub>CSi), 0.92 (9 H, s, Me<sub>3</sub>CSi), 2.17 (3 H, s, C<sub>2</sub>-CH<sub>3</sub>), 4.10 (1 H, m, H-5), 4.34 (1 H, br s, H-3), 10.14 (1 H, s, HCO). IR (CHCl<sub>3</sub>) 2910, 1675, 1460, 1255, 1080, 840 cm<sup>-1</sup>.

(3S,5R)-3,5-Bls[(tert-butyldlmethylsllyl)oxy]-1-ethynyl-2-methylcyclohex-1-ene (4b). To a mixture of zinc dust (290 mg, 4.4 mmol) and Ph<sub>3</sub>P (1.14 g, 4.35 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL) at room temperature was added CBr4 (1.3 g, 4.0 mmol) and the resulting mixture was stirred for 15 min. Pyridine (1.4 mL) was then added, followed by a solution of aldehyde 13d (280 mg, 0.73 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL). After stirring for 30 min, Et<sub>2</sub>O (20 mL) was added and the resulting suspension filtered through a short pad of silica gel  $[Et_2O (2 \times 25 mL)]$ . Concentration of the filtrate gave a crude which was purified by flash chromatography (hexanes) to afford, after concentration in vacuo and high vacuum drying, 368 mg of dibromide 14b (92 %, mp: 62-64 °C, yellowish solid). <sup>1</sup>H NMR  $\delta$  0.06 and 0.10 (12 H, two br s, two Me<sub>2</sub>Si), 0.88 and 0.90 (24 H, two s, two MegCSi),1.66 (3 H, br s, C<sub>2</sub>-CH<sub>3</sub>), 4.03-4.13 (1 H, m, H-5), 4.14 (1 H, br t, J= 3.2 Hz, H-3), 6.98 (1 H, s, vinyl H). IR (CCl4) 1070, 1250 cm<sup>-1</sup>. MS, m/z 542 (M<sup>+</sup>, 1.3), 540 (2.3), 538 (1.1), 527 (10), 525 (17), 523 (8), 483 (44), 408 (74), 352 (72) 327 (96), 271 (58) 246 (49), 189 (60) 147 (74) 133 (70) 111 (76), 69 (100). HRMS calcd for C2tH4002Si2Br2 540.0915, found 540.0920.

A solution of n-butyllithium in hexane (0.6 mL, 2.3 M, 1.38 mmol) was added dropwise to a solution of the above dibromide 14b in THF (6 mL) at -78 "C and the resulting deep red mixture was warmed to room temperature and quenched with a saturated aqueous solution of NH<sub>4</sub>CI (5 mL). The resulting mixture was extracted with Et<sub>2</sub>O (2 x 20 mL). The combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated in the rotary evaporator to give a residue which was purified by flash chromatography (80 % EtOAc/hexanes), affording, after concentration in vacuo and high vacuum drying, 230 mg of the enyne 4b. identical in all respects to the material obtained by bis-t-butylsilylation of 4a and t an authentic specimen obtained by Okarnura's procedure<sup>30</sup> (91 %, colourless oil). <sup>1</sup>H NMR  $\delta$  0.07 (6 H, s, Me<sub>2</sub>Si), 0.11 (6 H, s, Me<sub>2</sub>Si), 0.89 (9H, s, Me<sub>3</sub>CSi), 0.91 (9H, s, Me<sub>3</sub>CSi), 1.93 (3 H, br s, C<sub>2</sub>-CH<sub>3</sub>), 3.06 (1 H, s, sp-CH), 4.04-4.15 (1 H, m, H-5). 4.21 (1 **H,** 1, J=4 Hz, H-3). IR (CC14) 3310, 2085, 1260,835 cm-l; MS, m/z 380 (M+, 3) 365 (M+-CH3, 25) 323 (80) 248 (100), 192 (44), 165 (50), 115 (9), 73 (75). HRMS calcd for C<sub>21</sub>H<sub>40</sub>O<sub>2</sub>Si<sub>2</sub> 380.2567, found 380.2547.

De-A,B-cholest-8-en-8-yl Trlfluoromethanesulphonate (3a). Lithium di-isopropylamide (LDA) was prepared by the addition of di-isopropylamine (0.16 mL, 1.14 mrnol) to a solution of n-BuLi in hexane (0.4 mL. 2.56 M. 1.02 mrnol) and THF (1 mL) at -78 °C. After stirring for 10 min at -78 °C and 15 min at room temperature, the solution was again cook to -78 °C and Grundmann's ketone (214 mg, 0.81 mmol) in THF (2 mL) was added dropwise via a cannula. After stirring f 15 min. the enolate solution was warmed to room temperature over 2 h and then retooled to -78 "C. N-phenyltriflimide (320 mg, 0.9 mmol) was dissolved in THF (2 mL) and added to the enolate at - 78 "C. The reaction mixture was warmed to °C and stirred for 10 h. The resulting solution was poured into water and extracted with Et<sub>2</sub>O. The organic phase was drie (MgS04). filtered and concentrated in the rotary evaporator. The residue was purified by chromatography (hexanes) to afford, after concentration in vacuo and high vacuum drying, 300 mg of 3a (86 %, colourless oil). <sup>1</sup>H NMR  $\delta$  0.75 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.85 and 0.88 (6 H, two br s, C<sub>26,27</sub>-CH<sub>3</sub>), 0.93 (3 H, d, J= 6.4 Hz, C<sub>21</sub>-CH<sub>3</sub>), 5.57 (1 H, m, H-9).

De-A,B-22-lodo-23,24-dinorcholestan-8β-ol (6c). A mixture of tosylate 6b<sup>14</sup> (1 g, 2.7 mmol), acetone (50 mL, freshly purified with KMnO<sub>4</sub> and distilled from K<sub>2</sub>CO<sub>3</sub>) and Nal (1.5 g, 10 mmol) was stirred in the dark at 60-70 °C for 6 h. Water (10 mL) was added, the resulting solution was concentrated in the rotary evaporator to a small volume, and this residue was extracted with Et<sub>2</sub>O (3 x 45 mL). The combined organic extracts were washed with brine, dried (MgSO<sub>4</sub>), filtered and concentrated in the rotary evaporator to give a crude product which was purified by chromatography (5-10 % EtOAc/hexanes) to afford 0.84 g of the iodide 6c (96 %, viscous colourless liquid). <sup>1</sup>H NMR 8 0.97 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 1.00 (3 H, d, J= 5.5 Hz, C216H3), 3.18 (1 H, dd, J= 4.6 and9.5 Hz, CHHI), 3.32 (1 H, dd, J= 2.1 andg.5 Hz, CHHI), 4.09 (1 H, m, H-8). MS, m/z 322 (M+, 1), 307 (M+-CH3, 6), 177 (77), 135 (29), 111 (100). HRMS calcd for C13H23Ol: 322.0797, found 322.0794.

De-A,B-27-norcholestan-8ß-ol-25-one (7a). To a sonicated (150 W) solution of iodide 6c (140 mg, 0.44 mmol) and methyl vinyl ketone (65 µL, 0.8 mmol) in deoxygenated 7:3 EtOH/H<sub>2</sub>O (6 mL) were added Zn dust (110 mg, 1.7 mmol, freshly purified and dried<sup>36</sup>) and Cul (42 mg, 0.44 mmol, freshly purified and dried<sup>37</sup>). The mixture was sonicated under argon at room temperature for 20 min, and then additional amounts of Zn (55 mg, 0.85 mmol) and Cul (42 mg, 0.23 mmol) were added. After further sonication for 30 min, Et<sub>2</sub>O (10 mL) was added. The resulting mixture was filtered, and the solids were washed with Et<sub>2</sub>O (3 x 30 mL). The filtrate was washed with brine and the aqueous phase was extracted with Et<sub>2</sub>O (15 mL). The combined organic extracts were dried (MgSOq), filtered and concentrated in the rotary evaporator. Flash chromatography of the crude (5 % EtOAc/hexanes) afforded, after concentraton in vacua and high vacuum drying, 67 mg of the ketone 7a (76%), identical with an authentic sample (TLC,  $1H NMR$ ) obtained by an alternative route.17

De-A,B-25,25-(ethylenedloxy)-27-norcholest-8-en-8-yt trlfluoromethanesulphonate (3b). The same procedure as for the preparation of 3a was used to obtain 3b (90 %, colourless oil). <sup>1</sup>H NMR  $\delta$  0.75 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.93 (3 H, d, J= 6.7 Hz, C<sub>21</sub>-CH<sub>3</sub>), 1.32 (3 H, s, C<sub>26</sub>-CH<sub>3</sub>), 3.93 (4 H, m, OCH<sub>2</sub>CH<sub>2</sub>O), 5.57 (1 H, dd, J= 6 and 3.5 Hz, H-9); lR (CHCl3) 2940, 1410,1245, 1145,680 cm-l; MS, m/z 425 (M+-CH3,7), 361 (1) 87 (100) 69 (6) 43 (12); HRMS calcd for C2cH3105F3S -CH3 425.1609, found 425.1616.

6,7-Dehydro-1a-hydroxyprevitamin D<sub>3</sub> (2a). A mixture of enynediol 4a (32 mg, 0.211 mmol), triflate 3a (93 mg, 0.234 mmol), Et3N (0.115 mL, 0.805 mmol), bis-triphenylphosphine palladium dichloride (4 mg, 0.005 mmol) and DMF (2 mL) was heated at 75 °C for 10 h and then was cooled to room temperature. Et<sub>2</sub>O was added and the ether layer was washed successively with water and brine, dried (MgSO4) and filtered. Concentration in the rotary evaporator gave a residue which was purified by flash chromatography (40-50 % EtOAcIhexanes) to afford, after concentration in vacua and high vacuum drying, 45 mg of the dienyne 2a (54 %, viscous liquid); <sup>1</sup>H NMR 8 0.69 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 1.98 (3 H, t, J= 1.8 Hz, ClgCH3), 4684.13 (1 H, m, H-3) 4.24 (1 H, 1, J= 4 Hz, H-l), 5.97 (1 H, 1, J= 3.2 Hz, H-9); MS m/z 398 (M+, 2) 397 (6) 396 (28), 380 (M+-H<sub>2</sub>O, 8), 379 (28), 378 (100), 361 (8), 360 (30). HRMS calcd for C<sub>27</sub>H<sub>42</sub>O<sub>2</sub>-H<sub>2</sub>O 380.3079, found 380.3076.

1a-tert-Butyldimethylsllyloxy-6,7-dehydro-25,25-(ethylenedloxy)-27-norprevitamin D3 tertbutyldlmethylsllyl ether (2b). A mixture of enyne 4b (80 mg, 0.21 mmol), triflate 3b (90 mg, 0.20 mmol), Et<sub>3</sub>N (0.08 mL, 0.56 mmol), bis-triphenylphosphine palladium dichloride (4 mg, 0.005 mmol) and DMF (1.5 mL) was heated at 75 °C for 1.5 h and then cooled to room temperature. The reaction mixture was diluted with  $Et<sub>2</sub>O$  and the resulting mixture was washed with water and brine. Drying (MgSO<sub>4</sub>), filtration and concentration in the rotary evaporator gave a residue which was purified by flash chromatography (2-4 % Et2O/hexanes) to afford, after concentration in vacuo and high vacuum drying, 123 mg of the dienyne 2b (91 %, a viscous liquid that decomposes rapidly even at -10 "C but is stable in solution in the refrigerator). <sup>1</sup>H NMR  $\delta$  0.06 and 0.090 (12 H, two s, Me<sub>2</sub>Si), 0.69 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.88 and 0.89 (24 H, two s, Me<sub>3</sub>CSi), 1.32 (3 H, s, C<sub>26</sub>-CH<sub>3</sub>), 1.90 (3 H, s, C<sub>19</sub>-CH<sub>3</sub>), 3.94 (4 H, m, OCH<sub>2</sub>CH<sub>2</sub>O), 4.13 (1 H, m, H-3), 4.19 (1 H, t, J= 3.6 Hz, H-1), 5.97 (1 H, d, J= 3 Hz, H-9); MS, m/z 572 (48), 554 (M+-HSiMe<sub>2</sub>t-Bu, 100), 538 (64), 513 (19), 422 (23), 355 (57), 301 (35). HRMS calcd for C4oH7004Si2-HSiMe2f-Bu 554.3793, found 554.3782.

1a-ferf-Butyldimethylsilyloxy-25,25-(ethylenedioxy)-27-norprevitamin D3 ferf-butyldimethylsilyl ether (15). To a solution of dienyne 2b (100 mg, 0.15 mmol) in hexane (5 mL) was added 0.5 mL of a solution of quinoline in hexane (100 µL of quinoline in 5 mL of hexane). Lindlar's catalyst from Aldrich (60 mg, previously dried at 60 °C for 4 h in high vacuum) was added, and the resulting solution was exposed to hydrogen gas at slightly positive H<sub>2</sub> pressure. After stirring for 45 min, TLC (2 % EtOAc/hexanes) indicated that all the starting material had been converted to a product with a slightly higher Rf; it is prudent to follow the reaction by TLC in order to avoid over-hydrogenation. After filtration and concentration in the rotary evaporator, the residue was purified by flash chromatography (1-3 % Et2O/hexane) to give, after concentration in vacuo and high vacuum drying, 95 mg of protected previtamin 15 (95 %, colouriess oil). <sup>1</sup>H NMR  $\delta$  0.06 and 0.09 (12 H, two s, two Me<sub>2</sub>Si), 0.69 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.89 (18 H, br s, two Me<sub>3</sub>CSi). 0.95 (3 H, d, J= 6.4 Hz, C<sub>21</sub>-CH<sub>3</sub>),1.32 (3 H, s, C<sub>26</sub>-CH<sub>3</sub>), 1.65 (3 H, s, C<sub>19</sub>-CH<sub>3</sub>), 3.94 (4 H, m, OCH<sub>2</sub>CH<sub>2</sub>O), 4.11 (2 H, br s, H-1,3), 5.55 (1 H, br s, H-9), 5.72 and 5.88 (2 H, AB, J= 12.1 Hz, H-6,7); UV (Et2O)  $\lambda_{\text{max}}$  253 nm (ε 7800 nm).

1α-fert-Butyldimethylsilyloxy-25,25-(ethylenedioxy)-27-norvitamin D3 fert-butyldimethylsilyl ether (16). Previtamin 15 (75 mg, 0.11 mmol) was dissolved in dry iso-octane (5 mL) and heated under reflux in the dark for 4 h. Concentration in the rotary evaporator at room temperature gave a residue which was purified by flash chromatography (2-4 % Et2O/hexanes) to afford, after concentration in vacuo and high vacuum drying, 71 mg of an 88:12 mixture (NMR ratio) of 16 and 15 (94 % combined yield, 82 % of vitamin 16). The mixture was inmediately subjected to deprotection. <sup>1</sup>H NMR of the mixture: 8 0.06 (12 H, br s, two Me<sub>2</sub>Si), 0.52 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.87-0.91 (18 H, br s, two Me<sub>3</sub>CSi), 0.93 (3 H, d, J= 6 Hz, C<sub>21</sub>-CH<sub>3</sub>),1.32 (3 H, s, C<sub>26</sub>-CH<sub>3</sub>), 3.94 (4 H, m, OCH<sub>2</sub>CH<sub>2</sub>O), 4.17-4.23 (1 H, m, H-3), 4.36 (1 H, m, H-1), 4.86 (1 H, br s, Z - H-19), 5.17 (1 H, br s, E - H-19),6.01 and 6.24 (2 H, AB, J= 11.8 Hz, H-6,7); UV (Et2O) λmax263 nm (ε 12500 nm); MS, m/z 672 (M+, 20), 657 (M+-CH3, 10), 646 (6), 540 (24), 355 (15), 311 (20), 211 (22), 163 (29), 113 (35), 108 (35), 108 (40), 104 (100), 87 (52).

1a-Hydroxy-25-oxo-27-norvitamin D<sub>3</sub> (17). A solution of the above mixture of 15 and 16 (65 mg, 0.09 mmol) in MeOH (5 mL) and CH<sub>2</sub>Cl<sub>2</sub> (0.6 mL) was stirred with AG 50W-X4 resin (1.5 g, prewashed with MeOH) at room temperature for 12 h in the dark. An additional amount of resin (0.5 g) was added and the stirring continued for 12 h. Filtration. concentration in the rotary evaporator and purification of the crude product by flash chromatography (50-70 % EtOAc/hexanes) afforded, after concentration in vacuo and high vacuum drying, 30 mg of pure ketone 17 (81 % from the mixture, white syrup). <sup>1</sup>H NMR  $\delta$  0.54 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.94 (3 H, d, J= 6.2 Hz, C<sub>21</sub>-CH<sub>3</sub>), 2.14 (3 H, s, C<sub>26</sub>-CH<sub>3</sub>), 4.23 (1H, m, H-3), 4.44 (1H, t, J= 4.6 Hz, H-1), 5.01 (1H, br s, H-19 E), 5.32 (1H, m, H-19 Z), 6.01 and 6.37 (2H, AB, J= 11.3 Hz, H-6,7); UV (Et<sub>2</sub>O)  $\lambda$  max 264 nm ( $\varepsilon$  20000); IR (film) 3360 (br), 2900, 1690, 1340, 1040 cm<sup>-1</sup>; MS, m/z 382 (M<sup>+</sup>, 86), 145 (18), 131 (20), 87 (39), 55 (36), 43 (100); HRMS calcd for C<sub>26</sub>H<sub>38</sub>O<sub>2</sub> 382.2872, found 382.2874.

1a,25-Dihydroxyvitamin D<sub>3</sub> (1b). A solution of methyllithium in diethyl ether (0.33 mL,1.3 M, 0.43 mmol), was added dropwise to a cooled -78 °C solution of 17 (30 mg, 0.08 mmol) in diethyl ether (6 mL). After stirring for 20 min, the reaction was quenched with water (1 mL). Concentration in the rotary evaporator at room temperature (in the dark) gave a residue which was diluted with CHCl3 and transferred to a separating funnel. Water was added and the aqueous phase was extracted with CHCl<sub>3</sub> (3 x 20 mL). The combined organic extracts were dried (MgSO<sub>4</sub>), filtered and concentrated in the rotary evaporator as above. The resulting crude product was flash chromatographed (70 % EtOAc/hexane) to give, after concentration in vacuo and high vacuum drying, 25 mg (80 %) of  $1\alpha$ , 25-dihydroxyvitamin D<sub>3</sub> (1b). Crystallization from benzene/EtOAc afforded white crystals [mp 94-96 °C (Lit.<sup>7b</sup> mp 94-96 °C)]. <sup>1</sup>H NMR  $\delta$  0.54 (3 H, s, C<sub>18</sub>-CH<sub>3</sub>), 0.94 (3 H, d J= 6.2 Hz, C<sub>21</sub>-CH<sub>3</sub>), 1.26 (6 H, s, C<sub>26,27</sub>-CH<sub>3</sub>), 4.23 (1 H, m, H-3), 4.43 (1 H, m, H-1), 5.00 (1 H, br s, H-19 E), 5.33 (1 H, m, H-19  $Z$ ), 6.01 and 6.37 (2 H, AB, J= 12.4 Hz, H-6,7).

Acknowledgement. This research was supported by Doctoral Research Fellowships (FPI) to J.L.M. and L.A.S. and by Grants (CAICYT and DGICYT-PB87-0478) by the Spanish Ministetio de Educacidn y Cienda. We thank Hoffmann la Roche (Basel) for the generous gift of vitamin  $D_2$  and vitamin  $D_3$ 

#### References and notes

- 1. Dedicated to Prof I. Ribas on the Occasion of his 89th birthday.
- 2. For preliminary communications describing part of this work, see: (a) Castedo, L.; Mascarefias, J. L.; Mourino, A. *Tetrahedron Letr.* 1987, 29, 2099. (b) Castedo, L.; Mascarenas, J.L.; Mouriti, A.; Sarandeses, L. A. *Tetrahedron Lett.* **1988, 29, 1203. (c) Mouriño, A.; Castedo, L.; Fernández, B. R.; Granja, J.; Maestro, M. A.; Mascareñas, J. L.;** Sarandeses, L. A. Vifamih D. Molecular, Cellular *and Clinical Endocrinology* Norman, A. W., Schaefer. K.. Grigoleit, H.-G., Herrath, D. v., Eds.: Walter de Gruyfer & Co: Berlin-NY, 1988; pp 34-42.
- 3. (a) Norman, A. W. *Wamin 0, the Calcium Homeostatic Steroid* Hormone; Academic Press: New York. 1979. (b) DeLuca, H. F.; Paaren, H. G.: Schnoes, H. K. Top. Curr. *Chem.* 1979,83,1. (c) Jones, H.; Rasmusson, G. H. *Prog. Chem. Org. Nat.* Prod. 1980.39, 63. (d) Dickson, I. Nature 1987, 325, 18.
- 4. (a) Miyaura, C.; Abe, E.; Kurfbayashi, T.; Tanaka, H.; Konno, K.; Nishii, Y.: Suda, T. *Biochem. Biophys. Res. Commun.* 1981, 102,937. (b) Abe, E.; Miyaura, C.; Sakagami. H.; Takeda, M.; Konno, K.; Yamazaki, T.; Yoshiki. S.; Suda, T. *Proc. NaN. Acad. Sci. USA* 1981, 78, 4990. (c) Colston, K.; Colston, M. J.; Feldman, D. *Endocrinology 1981, 108,* 1083. (d) Ostrem, V. K.; DeLuca, H. F. Steroids 1987, 49, 73. (e) *vitamin D. Molecular, Cellular ano Clinical Endocrinology* Norman, A. W., Schaefer, K., Grigoleit, H.-G., Herrath, D. v., Eds.; Walter de Gruyter & Co: Berlin-NY, 1988; and ref. therein.
- 5. (a) Lythgoe, B. Chem. Sot. Rev. 1980, 9, 449. (b) Pardo, R.; Santelli, M. Bull. Sot. *Chim. Fr.* 1985. 98. (c) Quinkerl, G. Ed.; *Synform1985, 3.41;* Ibid, 1986,4, 131; Ibid, 1987, 5, 1.
- 6. (a) Semmler. F. J.; Holick, M. F.; Schnoes, H. K.; DeLuca, H. F. *Tetrahedron Lett. 1972, 4147.* (b) Rubio-Lightbourn, J.; Morfsaki, M.; Ikekawa, N. *Chem. Pharm. Bull. 1973, 21, 1854. (c)* Rubio-Lightboum, J.; Morisaki, M.; Ikekawa, N.; Takesita, T. *Chem. Pharm. Bull. 1973,21, 2588.* (d) Barton, D. H. R.; Hesse, R. H.; Pechet, M. M.; Rizzardo, E. J. *Chem. Sot., Chem. Commun. 1974, 203. (e)* Narwid, T. A.; Bount, J. F.; lacobellf, J. A.; Uskokovic, M. R. He/v. *Chim. Acra 1974, 57, 781.* (1) Sato, T.; Yamauchi, H.; Ogata, Y.; Tsujii, M.; Kunii, T.; Kagel, K.; Toyoshima, S.; Kobayashi, T. *Chem. Pharm. Bull. 1978, 26, 2933. (g)* Ochi, K.; Matsunaga, I.; Nagano, H.; Fukushima, M.; Shindo, M.; Kaneko, C.; Ishikawa, M.; DeLuca, H. F. J. *Chem. Sot., Perkin Trans. I 1979, 185.* (h) Fiirst, A.: Labler, L.; Meier, W. *He/v. Chim. Acta 1981, 64. 1870.* (i) Fiirst, A.; Labler. L.; Meier, W. */bid 1982, 65,*  1499.
- 7. (a) Paaren, H. E.; DeLuca, H. F.; Schnoes, H. K. *J. Org. Chem.* 1980, 45,3253. (b) Vanmaele, L.; De Clerq, P. J.; Vandewalle, M. *Tetrahedron 1985, 41, 141.*
- 8. Andrews, D. R.; Barton, D. H. R.; Hesse, R. H.; Pechet, M. M. *J. Org.* Chem. 1988,51,4819.
- 9. (a) Baggiolini, E. G.; lacobelli, J. A.; Hennessy, B. M.; Uskokovic, M. R. *J. Am. Chem. Soc.* 1982, 104, 2945. (b) Baggiolini, E. G.; lacobelli, J. A.; Hennessy, B. M.; Batcho, A. D.; Sereno, J. F.; Uskokovic, M. R. *J. Org. Chem. 1986.51, 3098.*
- 10. (a) Harrison, R. G.; Lythgoe, 8.; Wright, P. W. *J. Chem. Sot. Pelkin Trans. I 1974, 2854.* (b) Dawson. T. M.; Dixon, J.; Littlewood, P. S.; Lythgoe, B.; Saksena, A. K. *J. Chem. Soc. (C)* 1971, 2960.
- 11. Castedo, L.; Mouriño, A.; Sarandeses, L. A. *Tetrahedron Lett.* 1986, 27, 1523.
- 12. Large-scale preparation of this ketone is now carried out in our laboratory by ozonolysis of vitamin D3 in methanolpyridine, followed by in situ sodium borohydride reduction and oxidation of the resulting alcohol with pyridinium dichromate.
- 13. (a) Hendrickson, J. B.; Bergeron, R. *Tetrahedron Lett.* **1973**, 4607. (b) McMurry, J. E.; Scott, W.J. *Tetrahedror Lert.* **1963,** 24, 979.
- 14. Sardina, F. J.; Mouriño, A.; Castedo, L. J. Org. Chem. 1986, 51, 1264.
- 15. Petrier, C.; Dupuy, C.: Luche, J. L. *Tetrahedron Left* **1986,** 27, 3149.
- 16. This finding allows the previous synthesis of 7a to be shortened by several steps.<sup>17</sup>
- 17 Mascareñas, J. L.; Mouriño, A.; Castedo, L. J. Org. Chem. **1986**, 51, 1269.
- 16. Luche, J. L. *J. Am. Chem. Sot.* **1978, 100,** 2226.
- 19. (a) Kogure, I.; Ojima, I. *J. Organometal. Chem.* 1982, 234, 249. (b) Schroeter, S. H.; Eliel, E. L. *J. Org. Chem.*  1965, 30, 1. (c) Yasuda, A.; Yamamoto, H.; Nozaki, H. &II/. Chem. Sot. *Japan* **1979,** 52, 1757. (d) McChesney, J. D.; Thomson, T. N. *J. Org. Chem.* **1986,50,3473.**
- 20. Mitsunobu, I. *Synthesis* 1981, 1.
- 21. (a) Klein, E.; Ohloff, G. *Tetrahedron* **1963, 19,** 1091.
- 22. Tanabe, M.; Hayashi, K. *J. Am. Chem. Sot.* **1980,** 102,862.
- 23. Seyferth, D.; Andrews. S. B. *J. Organometal. Chem.* **1971, 30,** 151.
- 24. Still, W. C.; Mitra, A. *J. Am. Chem. Sot.* **1978, TOO, 1927.**
- 25. Sharpless, K. B.; Michaelson, R. C. *J. Am. Chem. Sot.* **1973,95,6136.**
- 26. The use of Mo(CO)6 as catalyst instead of VO(acac)<sub>2</sub> afforded less satisfactory results.
- 27. In this case,  $Mo(CO)6$  turned out to be superior to  $VO(acac)6$  as a catalyst for the transformation 11b-12b
- 28. Marcuso, A. J.; Browntain, D. S.; Swern, D. *J. Org. Chem.* **1979, 44,** 4148.
- 29. Corey, E. J.; Fuchs, P. L. *Tetrahedron Left.* **1972, 3769.**
- 30. While this work was in progress an excellent synthesis of this compound has appeared: (a) Aurrecoechea, J. M.; Okamura, W. H. *Tetrahedron Left.* **1987,28,** 4947. (b) Okamura, W.H.; Aurrecoechea, J.M.; Gibbs, R.A.; Norman, A.W. *J. Org. Chem.* **1989.** *54, 4073.*
- 31. Baggiolini, E. G.; Hennessy, B. M.; lacobelli, J. A.; Uskokovic, M. R. *Tefrahedron Left.* **1987,** *28, 2095.*
- 32. Kociensky, P. J.; Lythgoe, B. *J. Chem. Sot., Perkin Trans.* I **1980,** 400.
- 33. The use of the correspondig A-ring diacetate fragment also afforded moderate yield of dienyne, presumably due to the sensitivity of the allyl acetate to the palladium catalyst.
- 34. Scott, W. J.; Peña, M. R.; Swärd, K.; Stoessel, S. J.; Stille, J. K. *J. Org. Chem.* **1985**, *50, 2302.*
- 35. Still, W. C.; Kahn, M.; Mitra, A. *J. Org. Chem.* 1978, 43, *2923.*
- 36. Fieser, L. F.; Fieser, M. Reagents for *Organic Synthesis* John Wiley & Sons: 1967; vol 1, p 1292
- 37. Kauffman, G.B.; Teler, L.A. lnofg. *Synth.* **1963,** *7,9*